

Antimicrobial resistance of *Salmonella* Typhi and Paratyphi isolates in Portugal, January 1994 – June 2025

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1 Introduction

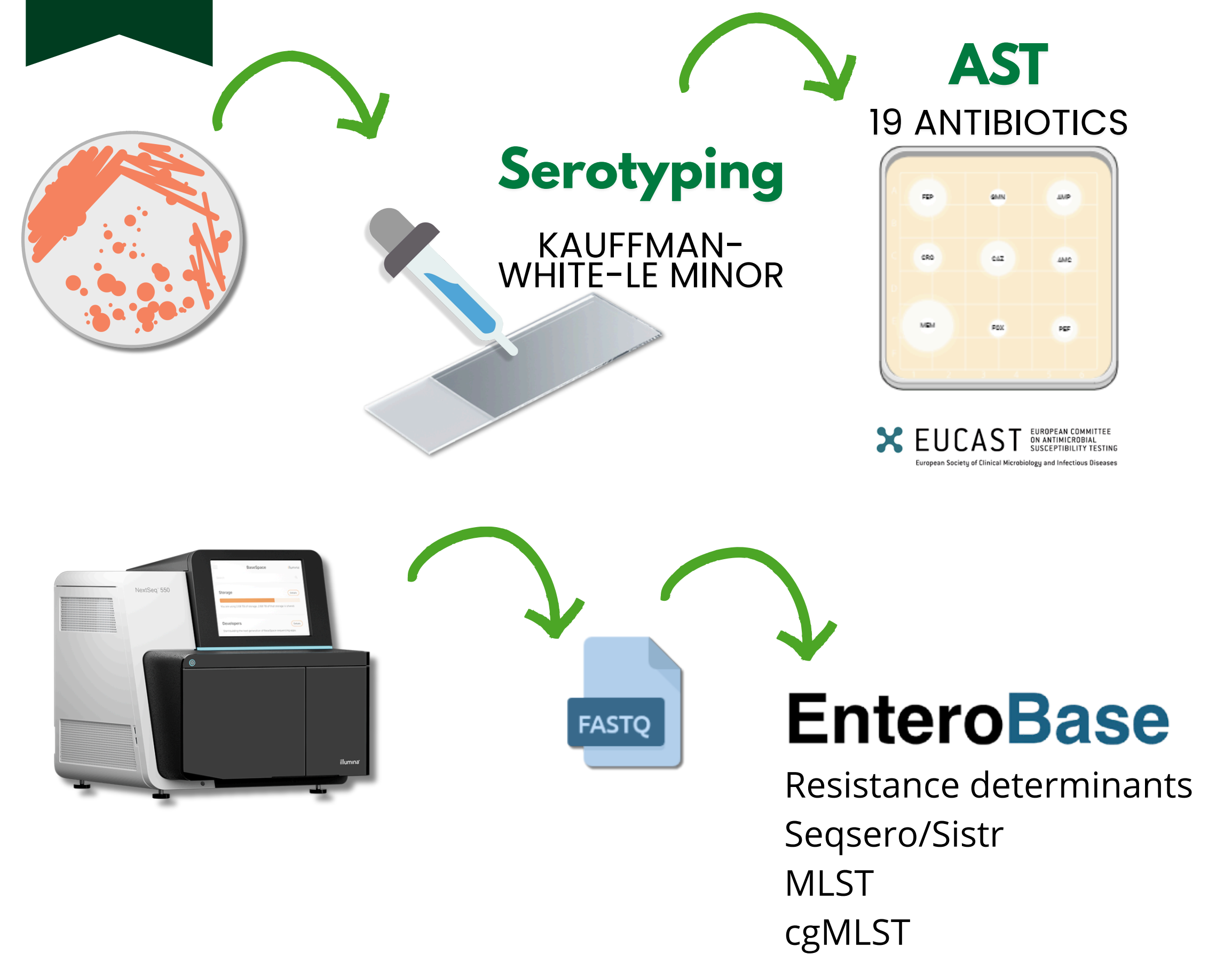
Typhoid and paratyphoid fever are severe systemic infections caused by *Salmonella* (*S.*) *enterica* subspecies *enterica* serovars Typhi and Paratyphi A, B and C.

Transmission occurs primarily through ingestion of food or water contaminated with faeces from infected individuals or via direct person-to person contact. In Europe, enteric fever is rare and mainly associated with travel to endemic countries.

The increasing prevalence of multidrug resistance (MDR), particularly in *S.* Typhi, poses significant therapeutic challenges.

Characterization of antibiotic resistance profiles of *S.* Typhi and Paratyphi A, B, and C isolates received in Jan 94-Jun 25

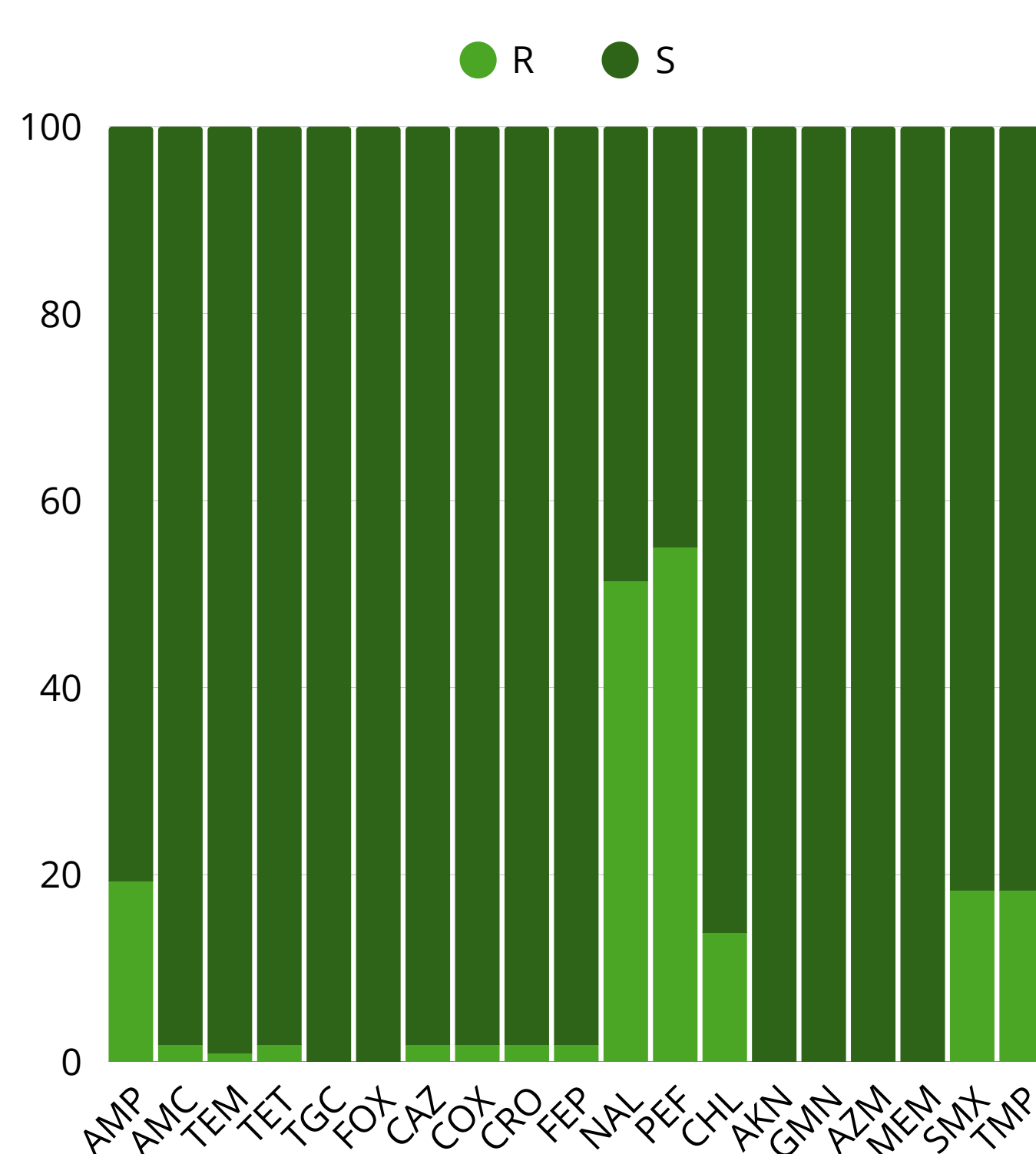
2 Methodology



3 Results

Total number of **191** isolates

- 167 Typhi
- 13 Paratyphi A
- 7 Paratyphi B
- 4 Paratyphi C



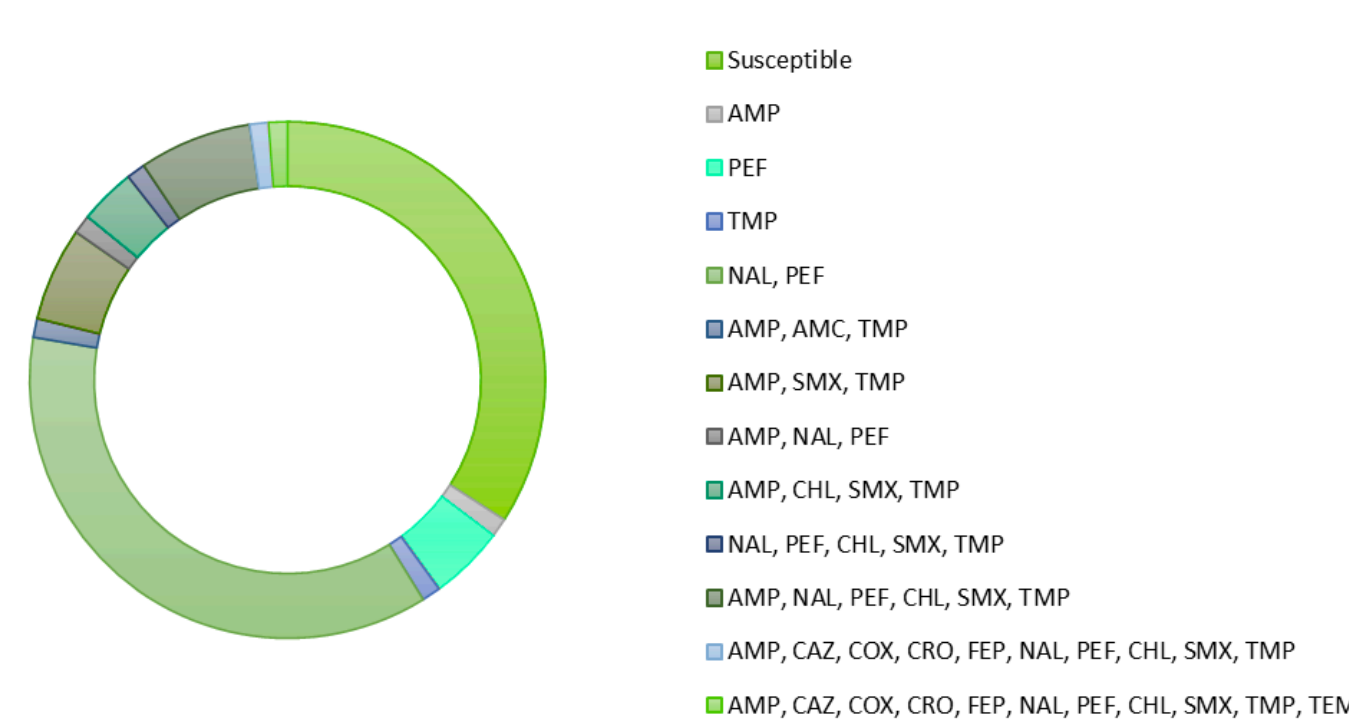
AMP: Ampicillin; AMC: Amoxicillin+Clavulanic acid; TEM: Temocillin; TET: Tetracycline; TGC: Tigecycline; FOX: Cefoxitin; CAZ: Ceftazidime; COX: Cefotaxime; CRO: Ceftriaxone; FEP: Cefepime; NAL: Nalidixic Acid; PEF: Pefloxacin; CHL: Chloramphenicol; AKN: Amikacin; GMN: Gentamicin; AZM: Azithromycin; MEM: Meropenem; SMX: Sulphamethoxazole; TMP: Trimethoprim.

Table 1. Phenotypic and genotypic antimicrobial characterization of the 109 sequenced isolates.

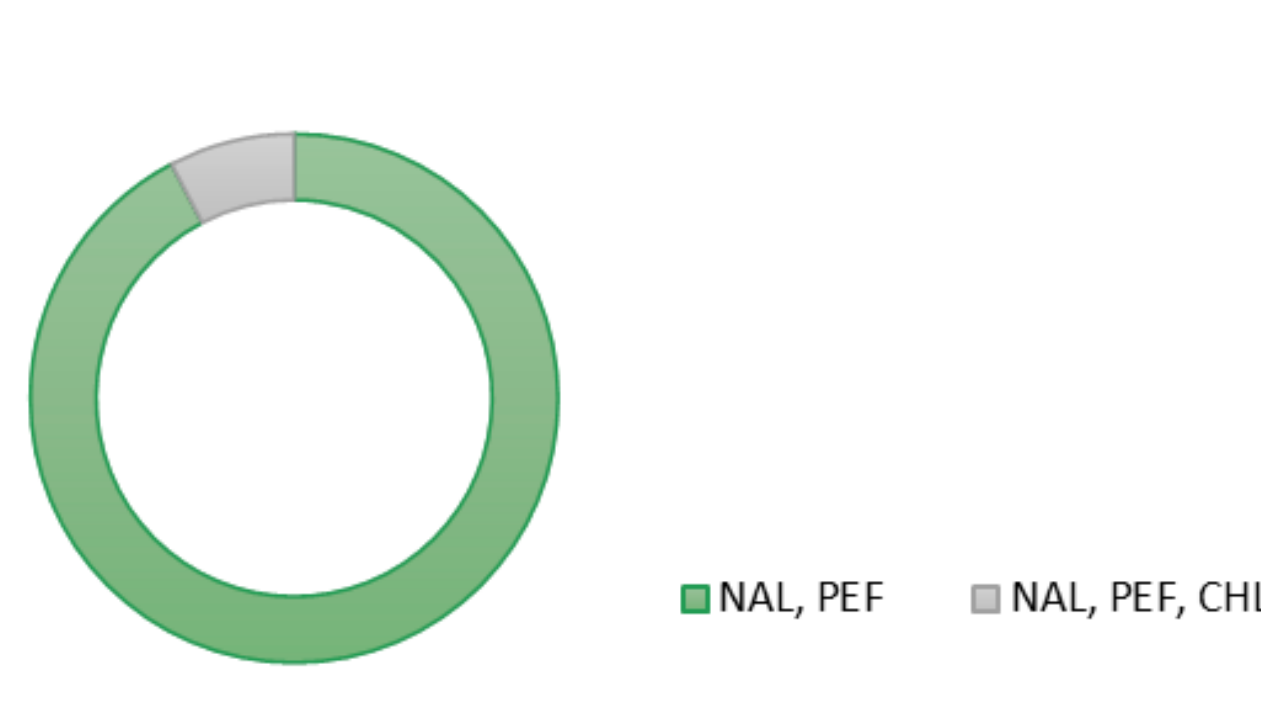
Phenotypic profile	Genotypic profile (resistance genes, point mutations, efflux pumps)	No. of isolates
S. Typhi (N=85)		
Susceptible	<i>aac(6)-laa</i>	28
	<i>aac(6)-laa, gyrB</i> (p.S464F)	1
AMP	<i>aac(6)-laa, blaTEM-1B</i>	1
PEF	<i>aac(6)-laa, gyrB</i> (p.S464F)	4
TMP	<i>aac(6)-laa, dfrA7</i>	1
	<i>aac(6)-laa, gyrA</i> (p.D87N)	2
	<i>aac(6)-laa, gyrA</i> (p.S83F)	22
	<i>aac(6)-laa, gyrA</i> (p.S83Y)	4
NAL, PEF	<i>aac(6)-laa, gyrA</i> (p.S83F, p.D87N)	1
	<i>aac(6)-laa, gyrA</i> (p.S83F, p.D87N), <i>parC</i> (p.S801)	1
	<i>aac(6)-laa, qnrB19, gyrA</i> (p.S83F)	1
AMP, AMC, TMP	<i>aac(6)-laa, blaTEM-1B, sul1, dfrA7</i>	1
AMP, NAL, PEF	<i>aac(6)-laa, blaTEM-1B, gyrA</i> (p.S83F)	1
AMP, SMX, TMP	<i>aac(6)-laa, blaTEM-1B, sul1, dfrA7</i>	5
AMP, CHL, SMX, TMP	<i>aac(6)-laa, aph(6)-Id, aph(3'')-Ib, blaTEM-1B, catA1, sul1, sul2, dfrA7</i>	3
NAL, PEF, CHL, SMX, TMP	<i>aac(6)-laa, gyrA</i> (p.S83F), <i>catA1, sul1, dfrA7</i>	1
AMP, NAL, PEF, CHL, SMX, TMP	<i>aac(6)-laa, aph(6)-Id, aph(3'')-Ib, blaTEM-1B, gyrA</i> (p.S83F), <i>catA1, sul1, sul2, dfrA7</i>	5
	<i>aac(6)-laa, aph(6)-Id, aph(3'')-Ib, blaTEM-1B, gyrA</i> (p.S83Y, p.D87G), <i>parC</i> (p.S801), <i>catA1, sul1, sul2, dfrA7</i>	1
AMP, CAZ, COX, CRO, FEP, NAL, PEF, CHL, SMX, TMP, TEM	<i>aac(6)-laa, aph(6)-Id, aph(3'')-Ib, blaTEM-1B, blaCTX-M-15*, catA1, qnrS1, gyrA</i> (p.S83F), <i>sul1, sul2, dfrA7</i>	1
AMP, CAZ, COX, CRO, FEP, NAL, PEF, CHL, SMX, TMP	<i>aac(6)-laa, aph(6)-Id, aph(3'')-Ib, blaTEM-1B, blaCTX-M-15*, catA1, qnrS1, gyrA</i> (p.S83F), <i>sul1, sul2, dfrA7</i>	1
S. Paratyphi A (N=13)		
	<i>aac(6)-laa, gyrA</i> (p.S83Y)	4
NAL, PEF	<i>aac(6)-laa, gyrA</i> (p.S83F), <i>parC</i> (p.T57S)	5
	<i>aac(6)-laa, gyrA</i> (p.S83F, p.D87N), <i>parC</i> (p.T57S)	1
	<i>aac(6)-laa, qnrB19, gyrA</i> (p.S83F)	2
NAL, PEF, CHL	<i>aac(6)-laa, gyrA</i> (p.S83Y), <i>parC</i> (p.T57S), <i>AcrAB-TolC</i>	1
S. Paratyphi B (N=7)		
Susceptible	<i>aac(6)-laa</i>	5
NAL, PEF	<i>aac(6)-laa, gyrA</i> (p.S83F)	1
AMP, AMC, TET, CHL, SMX	<i>aac(6)-laa, aadA2, blaCARB-2, tet(G), floR, sul1</i>	1
S. Paratyphi C / S. Choleraesuis (N=4)		
Susceptible	<i>aac(6)-laa</i>	3
AMP, TET, NAL, PEF, CHL, SMX, TMP	<i>aac(6)-laa, ant(3'')-Ia, aadA1, aadA2, blaTEM-1B, tet(A), qnrS1, cml1, sul3, dfrA12</i>	1

* Extended-spectrum beta-lactamase - ESBL; AMC, Amoxicillin-Clavulanic acid; AMP, Ampicillin; CAZ, Ceftazidime; CHL, Chloramphenicol; COX, Cefotaxime; CRO, Ceftriaxone; FEP, Cefepime; NAL, Nalidixic acid; PEF, Pefloxacin; SMX, Sulphamethoxazole; TEM, Temocillin; TET, Tetracycline; TMP, Trimethoprim.

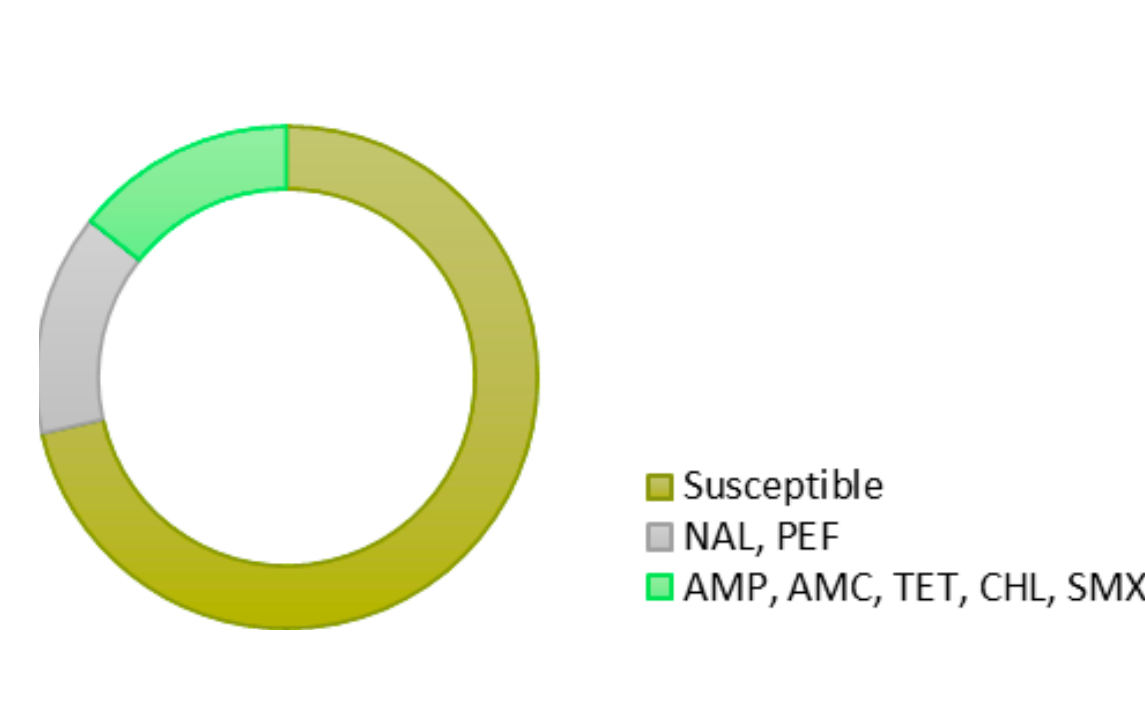
S. Typhi



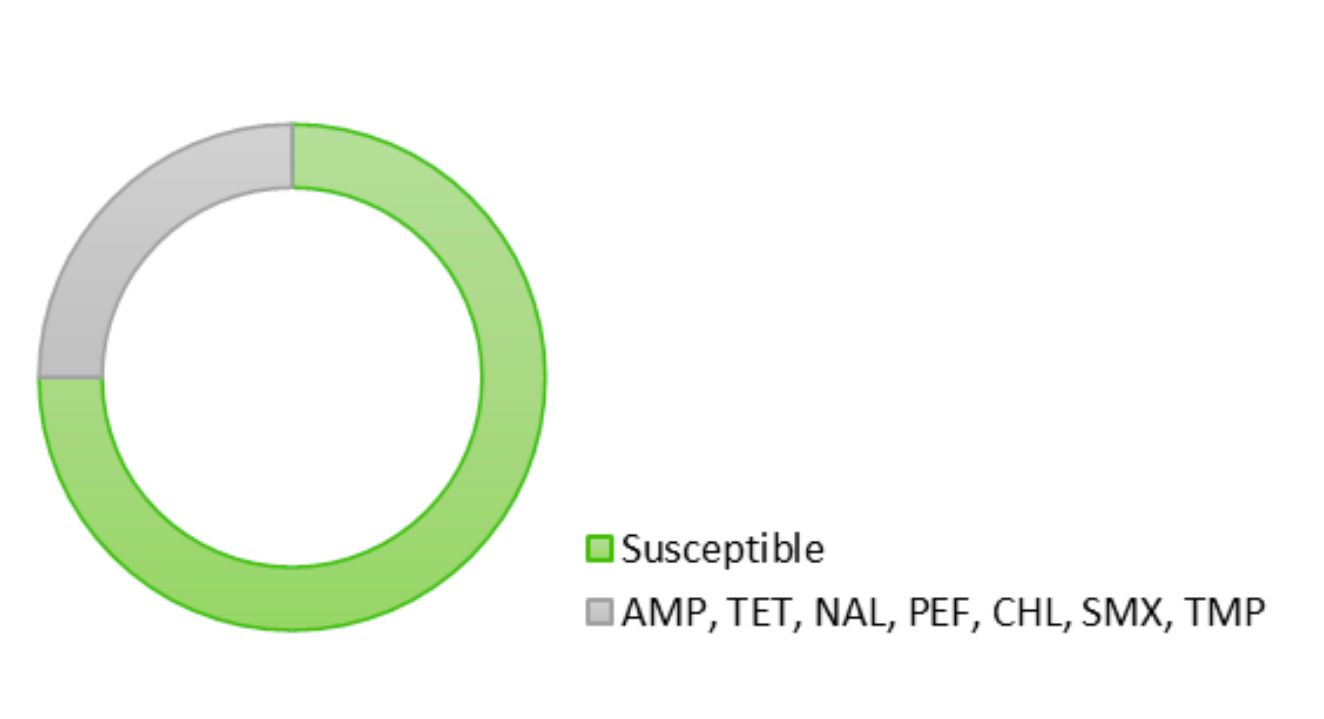
S. Paratyphi A



S. Paratyphi B



S. Paratyphi C



4 Conclusion

Although typhoid and paratyphoid serotypes are rare and non-endemic in Portugal, continued laboratory surveillance is essential to monitor circulating isolates, detect emerging resistance, support control measures and enable early identification of potential outbreaks.

